

25- 8-03;15:55

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s): James Harrison Aylward

Docket

: 14923Z

Serial No

: 09/888,178

Group Art Unit: 1654

Filed

: June 21, 2001

Examiner

: Christopher Robin Tate

For

: ANTI CANCER COMPOUNDS

Commissioner of

Patent and Trademarks

Washington, D.C. 20231

DECLARATION PURSUANT TO 37 C.F.R. §1.132

I, Dr. James Harrison Aylward, hereby declare as follows:

- I am currently the Research Director of Peplin Operations Pty Ltd, a subsidiary of Peplin Biotech Ltd, Ground Floor, South Tower, 527 Gregory Terrace, Bowen Hills, Brisbane, QLD, 4006, Australia. My Curriculum Vitae is attached hereto as Exhibit JHA-1.
- 2. I have published extensively in the area of biochemistry. A list of my publications is included in my Curriculum Vitae (Exhibit JHA-1).
- 3. I am an inventor of subject matter contained and described in United States Patent Application Serial No. 09/888,178 filed on 21 June, 2001 (hereinafter referred to as the "APPLICATION"). The APPLICATION is directed *inter alia* to a method for treating cancer by administering to the subject in need thereof a therapeutically effective amount of an angeloyl-substituted ingenane obtainable from the sap of a

Euphorbia species and an active derivative of an angeloyl-substituted ingenane obtainable from the sap of a Euphorbia species.

An example of a derivative of an angeloyl-substituted ingenane is an acetylated derivative. Acylation of the free accessible hydroxyls on ingenane 8 and 9 should improve their stability by preventing acyl migration. A number of acyl groups could be chosen, but as a test system, acetylation was selected. The chemical structure of ingenane 8 and 9 are shown in Exhibit JHA-2

- 4. Acyl migration is probably an intramolecular process involving attack of a free hydroxyl group on a closely situated carbonyl carbon. The developing positive charge on the attacking oxygen and the developing negative charge on the carbonyl oxygen are likely to be more stabilized in more polar solvents and thus this process is more likely to be observed in these. In the peplus milk the non-polar diterpenes and associated latex may form vesicles with non-polar interiors which act to protect the molecules from the polar aqueous environment, but these would be broken down on purification leaving the compounds more susceptible to such processes.
- 5. In conjunction with my scientific collaborators, I have conducted routine experiments in the acetylation of Ingenanes, the experimental details which follow acetylated derivatives of angeloyl—substituted ingenanes were tested for anticancer activity. All had strong bipolar activity of at least 1000 bipolar units, as measured by reversion of malignant melanoma MM96L cells to a bipolar dendritic morphology, the assay as described in United States Patent Application Serial No. 09/888,178 filed on 21 June, 2000.

Example 1:

Acetylation of a 6mg mixture of ingenane 8 and its 5-angeloyl isomer arising from acyl migration in ingenane 8, was carried out with acetyl chloride in dry pyridine to give a mixture which contained two major components identified as the 3-O-angelate-

5-O-acetate and the 5-O-angelate-3-O-acetate in quantities of 1.7 and 2.3mg (ca 50% combined yield). These had bipolar activity at about the same level as ingenane 8 and its 5-angeloyl isomer.

3-O-angelate-5-O-acetate

5-O-angelate-3-O-acetate

The experimental details of the acetylation of ingenane 8 are as follows. 3-angeloyloxy-4,5-dihydroxyingena-1,6-dien-9-one (6mg) was stood in methanol/water for several days at -4°C then concentrated, dissolved in anhydrous pyridine (100µl) and treated with acetyl chloride (10µl)at room temperature for 24h. Water (1ml) was added and the resulting mixture passed through a 7mm diam. x 20mm column of Chromatorex ODS resin (Fuji Silysia Chemical Co.). The resin was washed successively with water (7ml), 1:1 water:methanol (8ml) and methanol (36ml). The combined methanol eluates were concentrated and subjected to HPTLC on Merck 10 x 20cm HPTLC plate coated with LiChrospher Si60F_{254s} (eluent 50% MTB in 40-60° bp petroleum spirit). Concentration of the ether extract of the excised band with R_f 0.95 gave a colourless gum (4.4mg) which was subjected to HPLC on a 10mm diam. x 250mm Alltima C18-5u column with 68% methanol in water, isocratic for 40 minutes then a non-linear gradient to 100% methanol over 118 mins. Concentration of the eluate containing the peak at 165 mins. gave 5-acetoxy-3angeloyloxy-4-hydroxyingena-1,6-dien-9-one (compound I) as a colourless gum (2mg). APCIMS⁺ m/z 479 (7) [M+Na]⁺, 457 (4) [M+H]⁺, 397 (3) [M-OAc]⁺, 357 (8) [M-angelate]⁺, 315 (17) [M-angelate, -AcOH]⁺, 297 (100) [M-angelate, -AcOH, $-H_2O_1^+$, 269 (19) [M-angelate, $-H_2O_1$, $-CO_1^+$.

Concentration of the eluate containing the peak at 153 mins. gave a mixture which was resubjected to HPLC on a 10mm diam. x 250mm Alltima C18-5u column with 80% methanol in water, isocratic for 64 mins. then a non-linear gradient to 100% methanol over 21 mins. Concentration of the eluate containing the peak at 52 mins. gave 3-acetoxy-5-angeloyloxy-4-hydroxyingena-1,6-dien-9-one (compound II) as a colourless gum (2mg). APCIMS⁺ m/z 397 (6) [M-OAc]⁺, 315 (50) [M-angelate, -AcOH]⁺, 297 (100) [M-angelate, -AcOH, -H₂O]⁺, 269 (29) [M-angelate, -H₂O, -CO]⁺ (see Tables I, II and III for details).

Example 2:

Acetylation of a 10mg mixture of ingenane 9 and its isomers arising from acyl migration in ingenane 9, was carried out similarly to give a mixture which contained two major components identified as the 3-O-angelate-5,20-O-diacetate and the 5-O-angelate-3,20-O-diacetate in quantities of 0.9 and 1.2mg (ca 19% combined yield).

These had bipolar activity at about tenfold more than the ingenane 8 derivatives and it did not appear to vary significantly between the two isomers.

3-O-angelate-5,20-O-diacetate

5-O-angelate-3,20-O-diacetate

The experimental details of the acetylation of ingenane 9 are as follows.

3-angeloyloxy-4,5,20-trihydroxyingena-1,6-dien-9-one (10mg) was stood in methanol/water for several days at -4°C then concentrated, dissolved in anhydrous pyridine (200µl) and treated with acetyl chloride (20µl)at room temperature for 24h. Water (1ml) was added and the resulting mixture passed through a 7mm diam. x

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20mm column of Chromatorex ODS resin (Fuji Silysia Chemical Co.). The resin was washed successively with water (7ml), 1:1 water:methanol (8ml) and methanol (36ml). The combined methanol eluates were concentrated and subjected to HPTLC on Merck $10 \times 20 \text{cm}$ HPTLC plate coated with LiChrospher Si60F_{254s} (eluent 50% MTB in 40-60° bp petroleum spirit). Concentration of the ether extract of the excised band with R_f 0.89 gave a colourless gum (3.2mg) which was subjected to HPLC on a 10mm diam. x 250mm Alltima C18-5u column with 80% methanol in water, isocratic for 102 mins. then a linear gradient to 100% methanol over 14 mins.

Concentration of the eluate containing the peak at 52 mins. gave 3-angeloyloxy-5,20-bis(acetoxy)-4-hydroxyingena-1,6-dien-9-one (compound III) as a colourless gum (1mg). APCIMS⁺ m/z 537 (13) [M(C₂₉H₃₈O₈)+Na]⁺, 515 (3) [M+H]⁺, 313 (22) [M-angelate, -angelic acid, -CH₂CHO]⁺, 295 (100) [M-angelate, -angelic acid, -CH₂CHO, -H₂O]⁺.

Concentration of the eluate containing the peak at 30 mins. gave 5-angeloyloxy-3,20-bis(acetoxy)-4-hydroxyingena-1,6-dien-9-one (compound IV) as a colourless gum (1mg). APCIMS⁺ m/z 537 (13) [M(C₂₉H₃₈O₈)+Na]⁺, 497 (6) [M-OH]⁺, 455 (9) [M-AcO]⁺, 313 (19) [M-angelate, -angelic acid, -CH₂CHO]⁺, 295 (100) [M-angelate, -angelic acid, -CH₂CHO, -H₂O]⁺ (see Tables I, II and III for details).

Note that the lower yield in this case is probably due to the other isomers which were produced in small quantities but not isolated. In both cases the yields are probably significantly lower than might be expected on a larger scale due to the problems with handling such small quantities (see Tables I, II and III for details).

These acetylated compounds appeared to be more stable than ingenane 8 and ingenane 9.

Table I ¹H NMR data (CD₂Cl₂, 500MHz) for compounds I-IV

	ta (CD ₂ Cl ₂ , 500MHz) for compounds I-IV δ (ppm)					
Н	I	П	Ш	IV		
1	6.05 q	6.06 bs	6.05 q	6.07 bs		
3	5.01 bs	4.97 bs	5.06 s	4.99 bs		
5	5.21 bs	5.33 bs	5.37 bs	5.47 bs		
7	5.83 dq	5.83 dq	6.22 bd	6.21 bd		
8	4.17 bd	4.19 bd	4.24 bdd	4.25 bd		
11	2.51 ddq	2.51 ddq	2.54 ddq	2.55 ddq		
12	2.29 ddd	2.30 ddd	2.27 ddd	2.28 ddd		
12'	1.72 ddd	1.75 ddd	1.75 ddd	1.79 ddd		
13	0.67 ddd	0.68 ddd	0.71 ddd	0.72 ddd		
14	0.86 dd	0.87 dd	0.92 dd	0.92 dd		
16	1.06 s	1.08 s	1.07 s	1.09 s		
17	1.04 s	1.05 s	1.05 s	1.Q6 s		
18	0.95 d	0.96 d	0.97 d	0.98 d		
19	1.75 d	1.75 d	1.76 d	1.76 d		
20	1.55 s	1.53 s	4.57 bd	4.47 bd		
20'	1.55 5	1.555	4.19 d	4.20 d		
3-OAng 2'-Me	1.89 dq		1.89 dq			
3-OAng 3'	6.13 qq		6.14 qq			
3-OAng 4'	1.97 dq		1.97 dq			
5-OAng 2'-Me	1.57 44	1.99 dq	1 22.	1.97 dq		
5-OAng 3'	 	6.23 qq		6.24 qq		
5-OAng 4'		2.01 dq	-	2.01 dq		
3-OAig 4		2.02.04		2.09 s		
5-OAc	2.26 s		2.22 s			
20-OAc	2.203		1.98 s	1.94 s		
4-OH	3.31 bs	3.13 bs	3.36 bs			
4-On	3.51 08					
J 1,19	1	1	J (Hz)	1.4		
J 7,8	5	5	5	5		
	1.4	1.4	 			
J 7,20	12	12	12	12		
J 8,14 J 11,12	4	3	3	3		
	4	5	5	5		
J 11,12'	$+\frac{4}{7}$	7	7	7		
J 11,18	16	16	16	16		
J 12,12'	10	10	10	9		
J 12,13	7	6	6	7		
J 12',13		8	8	8		
J 13,14	8	- °	12	13		
J 20,20'	14	1	1.4	1.4		
OAng J2'-Me,3'	1.7		1.8	1.4		
OAng J2'-Me,4'	1.4	7	7	7		
OAng J3',4'	7					

Table II. 13 C NMR data (CD₂Cl₂, 125MHz) for compounds I-IV

	δ (ppm)					
С	I	II*	Ш	ĪV		
1	132.8	132.9	132.5	132.7		
2	135.9	136.2	136.3	135.8		
3	82.6	83.0	82.2	82.7		
4	86.3	86.3	86.4	86.4		
5	78.0	78.1	75.4	74.4		
6	135.1	135.0	134.0	132.7		
7	128.1	126.4	132.1	131.8		
8	43.9	43.9	44.2	43.6		
9	206.2	#	205.8	206.7		
10	72.4	72.4	72.6	72.6		
11	39.2	39.1	39.1	39.5		
12	31.4	31.4	31.5	31.5		
13	23.5	23.5	23.6	23.6		
14	23.7	23.8	23.5	23.4		
15	24.8	24.8	24.8	24.8		
16	15.8	15.8	15.8	15.8		
17	28.7	28.7	28.7	28.7		
18	17.1	17.2	17.2	17.3		
19	15.7	15.7	15.7	15.7		
20	21.5	21.7	66.3	66.6		
3-OAng 1'	169.4	7	169.4			
3-OAng 2'	126.7		127.9			
3-OAng 2'-Me	21.0		21.0			
3-OAng 3'	139.3	1	139.7			
3-OAng 4'	16.1		16.2			
5-OAng 1'		167.1		166.3		
5-OAng 2'		127.6		127.3		
5-OAng 2'-Me		20.8		20.7		
5-OAng 3'		141.1		142.1		
5-OAng 4'		16.3		16.3		
3-OAc 1'		172.7		171.9		
3-OAc 2'		21.5		21.5		
5-OAc 1'	170.5		170.5			
5-OAc 2'	21.2		21.2			
20-OAc 1'			170.2	170.0		
20-OAc 2'			21.0	21.1		

^{*} incomplete spectrum
unable to determine value

Table III: Structures of compounds I-IV

compound I

compound II

Indications are that these compounds as expected are more stable than ingenanes 8 and 9, though 3-acetoxy-5-angeloyloxy-4-hydroxyingena-1,6-dien-9-one still has some stability problems.

6. It is my considered scientific opinion that these data support the claim that cancer can be treated by administering to the subject in need thereof a therapeutically effective amount of an angeloyl-substituted ingenane obtainable from the sap of a Euphorbia species and an active derivative of an angeloyl-substituted ingenane obtainable from the sap of a Euphorbia species.

The undersigned declares further that all statements made herein are of his own knowledge, are true, and that all statements made on information and belief are believed to be true, and further that these statements were made with the knowledge that willful, false statements, and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code.

Date: August 22 2003

Dr. James Harrison Aylward

EXHIBIT JHA-1

CURRICULUM VITAE

JAMES HARRISON AYLWARD

Home Address:

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Present Work Location:

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Australia

Date of Birth:

July 1, 1948, Springvale, Victoria, Australia

Present Position:

Research Director

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Ground Floor, South Tower

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Phone: +617 3854 0980 Fax: +617 3854 0989 Mobile: +61419 710 808

Email: jim.aylward@peplin.com

Marital Status:

Married, no children

Formal Education:

1972-75

PhD (Biochemistry) Monash University,

Clayton, Victoria, Australia

1970-71

MSc qualifying (Biochemistry), Monash

University, Clayton, Victoria, Australia

1967-69:

BSc, majors in Chemistry & Biochemistry,

Monash University, Clayton, Victoria,

Australia

1966:

Matriculation, Huntingdale High School,

Huntingdale, Victoria, Australia

Professional Experience:

April 1998 – present time

Research Director, Peplin Biotech

of research relating direction commercialisation of novel small molecules with biological activity, with focus on anticancer activity. Co-founder of Peplin

Biotech in 1998

1992 - April 1998:

Principal Research Scientist CSIRO Division of Tropical Agriculture 306 Carmody Road, St. Lucia, QLD 4068, Australia

Project Leader 1993-95 (Biotechnology group)

Budget responsibility: AUD \$1.5m pa

improving the nutrition of ruminants by increasing the nutritive value of dietary fibre by manipulation of enzymes of fibre degradation in the rumen, using the tools of protein biochemistry and molecular biology

enzymes for use in the paper pulp industry

use of bacteria and yeasts as biocontrol agents for protection of fruits and vegetables from fungal spoilage

agents for use in opportunistic fungal infections and as immune system boosters

anti-cancer compounds which promote cellular differentiation

development of new functional foods

DNA incorporation into bacteria using sub micron gold particles

Senior Research Scientist/Principal

Research Scientist CSIRO Division of Tropical Production. Meiers Road. Animal Indooroopilly, QLD, 4068, Australia

vaccines against tick-borne diseases

1981-83

Research Scientist/Senior Research Scientist CSIRO Division of Tropical Crops and Pastures, Cunningham Laboratory, St, Lucia QLD, 4068, Australia

1984-91

nutritive value and toxicity testing of new dietary legumes (beans) for ruminants and monogastrics

1980-81

Senior Tutor

Department of Monash University, Biochemistry, Clayton, VIC, 3168, Australia

control of intermediary metabolism by fragments of growth hormone in muscle, adipose tissue and liver

1979-80

Research Associate

Department of Physiology

Howard Hughes Medical Institute

Vanderbilt University, Nashville, Tennessee,

USA

mechanism of insulin and adrenalin action on muscle glycogen synthase, a key enzyme in

control of carbohydrate metabolism

1976-78

Research Associate

Department of Biochemistry

University of Miami School of Medicine

Miami, Florida, USA

enzymology of phosphorylase phosphatase, a key enzyme in energy metabolism under

hormonal control

Publications

Patent applications (CSIRO owned)

Inventors: Aylward, J.H. and Stone. B.F. (1991) "Tick paralysis toxin" Australia 86784

Inventors: Aylward, J.H. and Orpin, C.G. (1992) "Biocontrol bacteria" Australia PL 0256

Inventors: Williamson, M.A. and Aylward, J.H. (1992) "Biocontrol agents for use in horticulture" Australia PL 8298

Inventors: Aylward, J.H., Riddles, P.W., and Wright, I.G. (1993) "Antigens and polypeptides derived from Babesia (12D3) antigen." Australia 640398

Inventors: Aylward, J.H. and Williamson, M.A. (1993) "Biocontrol agents for use in agricultural products" Australia PL 7721

Inventors: Xue, G-P., Gobius, K.S., Aylward, J.H., and Orpin, C.G. (1993) "Recombinant cellulases"

Inventors: Aylward, J.H., and Williamson, M.A. (1996) "Biocontrol agents in treatment of opportunistic infections" Australia PN 9072

Non-CSIRO owned

Inventor: Aylward, J.H. (1997) "Anti-cancer compounds" Australia Provisional PO 8640, PCT/AU98/00656 (transferred to Peplin Biotech Pty Ltd)

Papers and Book chapters

Aylward, J.H., Bornstein, J., Gould, M.K. and Hall, S. (1972) Effect of polypeptides derived from growth hormone on the oxidation of pyruvate. Israel Journal of Medical Science 8 864.

Aylward, J.H., Bornstein, J., Gould, M.K. and Hall, S. (1974) Inhibition of muscle pyruvate dehydrogenase by a polypeptide from growth hormone. *Biochemical Biophysical Research Communications* 59 57-62.

Aylward, J.H. (1976) The effect of In-G on pyruvate dehydrogenase and glycogen synthase. Ph.D. Thesis, Monash University, Clayton, Victoria Australia.

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Lee, E.Y.C., Mellgren, R.L., Aytward, J.H. and Killilea, S.D. (1978) Mammalian phosphorylase phosphatase. Biochemical Society Transactions 6 25-29. Killilea, S.D., Aytward, J.H., Mellgren, R.L. and Lee, E.Y.C. (1978) Purification and properties of bovine mycardial

Killilea, S.D., Aylward, J.H., Mellgren, R.L. and Lee, E.Y.C. (1978) Purification and properties of bovine mycardia phosphorylase phosphatase (protein phosphatase C). Archives of Biochemistry and Biophysics 191 638-646.

Lee, E.Y.C., Mellgren, R.L., Killilea, S.D. and **Aylward, J.H.**(1978) Properties and regulation of liver protein phosphatases. In "Regulatory mechanisms of carbohydrate metabolism" (Ed V. Esmann) FEBS Symposium **42** 327-346 (Pergamon Press, New York).

Lee, E.Y.C., Aytward, J.H., Meligren, R.L., and Killilea, S.D. (1979) Protein phosphatase C: properties, specificity and structural relationship to a larger holoenzyme. In: "From gene to protein: Information transfer in normal and abnormal cells" (Eds. T.R. Russell et al), pp. 483-500 (Academic Press, new York).

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EXHIBIT JHA-2

Ingenane 8 (PEP006)

Ingenane 9 (PEP008)